

Result Interpretation

Semi-Quantitative Interpretation: Semi-quantitative results can be derived by simple comparison of the sample absorbances to the absorbance of the calibrators:

- Samples with a lower absorbance (less color) than a calibrator have a concentration of Aflatoxin greater than the concentration of the calibrator.
- Samples with a higher absorbance (more color) than a calibrator have a concentration less than the concentration of the calibrator.

Quantitative Interpretation: It is preferred for quantitative results to be determined using commercially available software for ELISA evaluation using a 4-parameter curve fit. Alternatively, a semi-log curve fit can be used if 4-parameter software is not available. A spreadsheet that will perform the curve fit and sample concentration calculations is available upon request. Please contact Beacon for further details.

- The concentration of Aflatoxin in a sample is determined by comparing the average sample absorbance to the standard curve. This value must then be multiplied by the dilution factor used.
- Samples with absorbances lower than the highest calibrator contain a concentration of Aflatoxin too high for quantification. Further dilute the sample extract in 16% methanol to fit into the standard curve and retest along with the calibrators. Results must then be multiplied by the dilution factor used.
- Samples with Aflatoxin absorbances greater than the lowest calibrator or less than the highest calibrator must be reported as < 2 ppb or > 100 ppb, respectively.

Technical Assistance

For questions regarding this kit or for additional information about Beacon products, contact us.

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Safety

Stop Solution is 1N hydrochloric acid. Handle with care. To receive complete safety information on this product, contact Beacon Analytical Systems, Inc., and request Safety Data Sheets.

General Limited Warranty

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Intended Use

The Beacon Aflatoxin 100 Tube Kit is an immunoassay for the detection of Aflatoxin in nuts, grain, and grain product samples. This product is intended for research use only.

Principles

Aflatoxin HRP Enzyme Conjugate is pipetted into the test tubes followed by the Calibrators and the Sample Extract(s). A soluble polyclonal Aflatoxin antibody solution is then added to the test tubes to initiate the reaction. During an incubation, Aflatoxin and Aflatoxin HRP Enzyme Conjugate compete for binding to the soluble Aflatoxin antibody which is in turn immobilized on the test tubes. Following the incubation, the tubes are washed to remove any unbound Aflatoxin and Aflatoxin HRP Enzyme Conjugate. After washing, a colorless substrate is added to the tubes and any bound enzyme conjugate will convert the substrate to a blue color. Following an incubation, the reaction is stopped with the addition of Stop Solution and the amount of color in each tube is measured. The color of the unknown sample is compared to the color of the calibrators and the Aflatoxin concentration of the sample is derived.

Reagents and Materials Provided

- 5 Bags each containing 20 test tubes that are vacuum sealed in an aluminized pouch with a desiccant.
- 5 Vials of Aflatoxin Calibrators (0, 2, 7.5, 25, and 100 ppb).
Note: The calibrators actually contain 1/10th of the stated value to account for the 1:10 dilution during sample preparation. No further correction is required to obtain the concentration of Aflatoxin in the sample.
- 1 Bottle of Aflatoxin HRP Enzyme Conjugate.
- 1 Bottle of Aflatoxin Antibody.
- 1 Bottle of Substrate.
- 1 Bottle of Stop Solution.

Reagents and Materials Required but Not Provided

- Pipette(s) with disposable tips capable of dispensing the required volume(s).
- Repeater pipette(s) with disposable tips capable of dispensing the required volume(s) (recommended if running more than five tubes at once).
- Laboratory quality distilled or deionized water.
- Reagents and materials for sample preparation.
- Personal protective equipment.
- Paper towels or equivalent absorbent material.
- Wash bottle (optional).
- Permanent Marker.
- Tube rack.
- Timer.
- Photometer capable of reading absorbance at 450 nm in 12 mm x 75 mm tubes.

Kit Handling Notes and Precautions

- Read the product brochure in its entirety prior to use.
- The kit, in its original packaging, can be used until the end of the month indicated on the box label.
- Do not use reagents after expiration date.
- Store all kit components at 4°C to 8°C (39°F to 46°F) when not in use.
- Reagents should be brought to room temperature, 20°C to 28°C (62°C to 82°F), prior to use. Avoid prolonged (> 24 hours) storage at room temperature.
- Do not freeze kit components or expose them to temperatures greater than 37°C (99°F).
- Running Calibrators and Samples in duplicate will improve assay precision and accuracy.
- Precise transfer of samples and reagents by using a calibrated pipette that is capable of dispensing the required volume is critical to obtain proper assay results.
- If running more than five tubes at once, the use of a repeater pipette is recommended when adding the Antibody, Substrate and Stop Solution.
- All procedural steps should be completed without interruption. Ensure all reagents, materials and equipment are ready at the appropriate time.
- Each reagent is optimized for use in the Beacon Aflatoxin 100 Tube Kit. Do not substitute reagents from any other manufacturer into the test kit. Do not combine reagents from other Beacon Aflatoxin 100 Tube Kits with different lot numbers.
- Dilution or adulteration of reagents or samples not called for in the procedure may result in inaccurate results.
- Damage to or obstruction of the optical surface may cause unsatisfactory results.

Specificity

The Beacon Aflatoxin 100 Tube Kit cannot differentiate between the various Aflatoxins but detects their presence to differing degrees. The following table shows the relative values for 50% B₀ and the percent cross-reactivity versus Aflatoxin B1. All concentrations are in ppb.

Compound	50% B ₀	% Cross-Reactivity
Aflatoxin B2	39	25
Aflatoxin G1	39.5	25
Aflatoxin G2	221	4

Extraction Solution Preparation (80% Methanol)

1. Measure 20 mL of distilled or deionized water for each 100 mL being prepared and transfer to a clean glass container with a tight-fitting lid.
2. Measure 80 mL of methanol for each 100 mL being prepared and add to the container.
3. Cover and swirl to mix. Store tightly sealed to minimize evaporative loss.

Sample Preparation

Corn and Other Grains:

1. Grind samples to pass through a 20 mesh sieve and thoroughly mix prior to sub-sampling. Samples not being immediately analyzed should be stored refrigerated.
2. Weigh 50 g of the ground sample and transfer to a clean blender jar.
3. Weigh 5 g NaCl and transfer to the blender jar.
4. Measure 100 mL of Extraction Solution and transfer to the blender jar.
5. Blend for 1 minute in a high-speed blender.
6. Filter a minimum of 10 mL through a paper filter (a coffee filter is recommended).
7. Dilute 5 mL of the extract with 20 mL of water and mix thoroughly.
8. Filter the diluted extract through a glass fiber filter.

Peanut Paste:

1. Weigh 50 g of the sample and transfer to a clean blender jar.
2. Measure 100 mL of Extraction Solution and add it to the jar.
3. Blend for 1 minute in a high-speed blender.
4. Filter a minimum of 10 mL through a paper filter (a coffee filter is recommended).
5. Dilute 5 mL of the extract with 20 mL of water and mix thoroughly.

Assay Procedure

1. Allow kit components and the sample extract(s) to reach room temperature prior to running the test.
2. Place the appropriate number of test tubes into a tube rack. Label the tubes one inch from the top with the calibrator concentration or sample identification. Be sure to re-seal unused tubes in the zip-lock bag with the desiccant to limit exposure to moisture.
3. Dispense **500 µL of Enzyme Conjugate** into each tube.
4. Dispense **500 µL of Calibrators and Sample Extract(s)** into the appropriate tube. Be sure to use a clean pipette tip for each solution to avoid cross contamination.
5. Dispense **500 µL of Antibody** into each tube.
6. Gently shake the tubes for 30 seconds using a back-and-forth motion and incubate for **10 minutes** at room temperature.
7. Decant the contents of the tubes into an appropriate waste container. Fill the tubes to overflowing with laboratory quality distilled or deionized water then decant. Repeat this wash step three times for a total of four washes. Following the last wash, tap the inverted tubes onto absorbent paper to remove excess wash solution.
8. Dispense **500 µL of Substrate** into each tube.
9. Gently shake the tubes for 30 seconds using a back-and-forth motion and incubate for **10 minutes** at room temperature.
10. Dispense **500 µL of Stop Solution** into each tube in the same order of addition as the Substrate.
11. Gently shake the tubes for 30 seconds using a back-and-forth motion.
12. Carefully wipe the optical surface with a soft, lint-free wipe. Measure and record the absorbance (Optical Density; OD) of each tube at 450 nm using a tube reader within 10 minutes of stopping the assay. Be sure to blank the reader with laboratory quality distilled or deionized water prior to measuring.