
Intended Use

The Beacon Enrofloxacin/Ciprofloxacin Plate Kit is an immunoassay for the detection of E/C in meat, fish, shrimp, and honey samples. This product is intended for research use only.

Principles

E/C HRP Enzyme Conjugate is pipetted into the test wells followed by the Calibrators and the Sample Extract(s). A soluble polyclonal E/C antibody solution is then added to the test wells to initiate the reaction. During an incubation, E/C and E/C HRP Enzyme Conjugate compete for binding to the soluble E/C antibody which is in turn immobilized on the test wells. Following the incubation, the wells are washed to remove any unbound E/C and E/C HRP Enzyme Conjugate. After washing, a colorless substrate is added to the wells and any bound enzyme conjugate will convert the substrate to a blue color. Following an incubation, the reaction is stopped with the addition of Stop Solution and the amount of color in each well is measured. The color of the unknown sample is compared to the color of the calibrators and the E/C concentration of the sample is derived.

Reagents and Materials Provided

- 1 Plate containing 12 test strips of 8 wells each that are vacuum sealed in an aluminized pouch with a desiccant.
- 6 Vials of E/C Calibrators (0, 0.1, 0.3, 0.6, 1.5, and 5 ppb).
- 1 Bottle of E/C HRP Enzyme Conjugate.
- 1 Bottle of E/C Antibody.
- 1 Bottle of Substrate.
- 1 Bottle of Stop Solution.

Reagents and Materials Required but Not Provided

- Pipette(s) with disposable tips capable of dispensing the required volume(s).
- Multichannel pipette(s) (8 channels) with disposable tips capable of dispensing the required volume(s) (recommended when running more than two strips at once).
- Laboratory quality distilled or deionized water.
- Reagents and materials for sample preparation.
- Personal protective equipment.
- Paper towels or equivalent absorbent material.
- Wash bottle (optional).
- Timer.
- Microtiter plate or strip reader capable of reading at 450 nm.

Kit Handling Notes and Precautions

- Read the product brochure in its entirety prior to use.
- The kit, in its original packaging, can be used until the end of the month indicated on the box label.
- Do not use reagents after expiration date.
- Store all kit components at 4°C to 8°C (39°F to 46°F) when not in use.
- Reagents should be brought to room temperature, 20°C to 28°C (62°C to 82°F), prior to use. Avoid prolonged (> 24 hours) storage at room temperature.
- Do not freeze kit components or expose them to temperatures greater than 37°C (99°F).
- Running Calibrators and Samples in duplicate will improve assay precision and accuracy.
- Precise transfer of samples and reagents by using a calibrated pipette that is capable of dispensing the required volume is critical to obtain proper assay results.
- If running more than two strips at once, the use of a multi-channel pipette is recommended when adding the Antibody, Substrate, and Stop Solution.
- All procedural steps should be completed without interruption. Ensure all reagents, materials and equipment are ready at the appropriate time.
- Each reagent is optimized for use in the Beacon Enrofloxacin/Ciprofloxacin Plate Kit. Do not substitute reagents from any other manufacturer into the test kit. Do not combine reagents from other Beacon Enrofloxacin/Ciprofloxacin Plate Kits with different lot numbers.
- Dilution or adulteration of reagents or samples not called for in the procedure may result in inaccurate results.
- Damage to or obstruction of the optical surface may cause unsatisfactory results.

Specificity

The Beacon Enrofloxacin/Ciprofloxacin Plate Kit cannot differentiate between the various Fluoroquinolones but detects their presence to differing degrees. The following table shows the relative values for the percent cross reactivity versus Enrofloxacin.

Compound	% Cross-Reactivity
Enrofloxacin	100
Ciprofloxacin	102
Danofloxacin	105
Norfloxacin	4

Limit of Detection (LOD)

Matrix	LOD (ppb)
Meat	High Method: 2, Low Method: 0.2
Fish	High Method: 2, Low Method: 0.2
Shrimp	High Method: 2, Low Method: 0.2
Honey	2

Sample Buffer 1 (40% Methanol)

1. Measure 60 mL of distilled or deionized water for each 100 mL being prepared and transfer to a clean glass container with a tight-fitting lid.
2. Measure 40 mL of methanol for each 100 mL being prepared and add to the container.
3. Cover and swirl to mix. Store tightly sealed to minimize evaporative loss.

Sample Buffer 2 (60% Methanol)

1. Measure 40 mL of distilled or deionized water for each 100 mL being prepared and transfer to a clean glass container with a tight-fitting lid.
2. Measure 60 mL of methanol for each 100 mL being prepared and add to the container.
3. Cover and swirl to mix. Store tightly sealed to minimize evaporative loss.

Sample Buffer 3 (10 mM PBS)

1. Measure 1 L of laboratory quality distilled or deionized water and add to a clean container with a tight fitting lid.
2. Weigh 0.31 g of $\text{NaH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$ and add to the container.
3. Weigh 2.87 g of $\text{Na}_2\text{HPO}_4 \cdot 12\text{H}_2\text{O}$ and add to the container.
4. Weigh 9 g of NaCl and add to the container.
5. Gently stir to mix.
6. Measure the pH of the solution and adjust to achieve a pH of 7.4, as necessary.

Sample Preparation

Meat, Fish, and Shrimp: High Detection Limit Method (Dilution Factor: 20)

1. Homogenize the sample.
2. Weigh 1 gram of sample into a 15 mL conical tube.
3. Measure 3 mL of Sample Buffer 1 and add to the tube.
4. Mix vigorously for 5 minutes.
5. Centrifuge for 10 minutes at 3,000 x g.
6. Transfer 0.4 mL of the supernatant to a clean 15 mL conical tube.
7. Measure 1.6 mL of Sample Buffer 3 and add to the tube.
8. Measure 2 mL of hexane and add to the tube.
9. Mix vigorously for 5 minutes.
10. Centrifuge for 10 minutes at 3,000 x g.
11. Transfer the lower phase to a clean vial and use in the assay.

Meat, Fish, and Shrimp: Low Detection Limit Method (Dilution Factor: 2)

1. Homogenize the sample.
2. Weigh 5 grams of sample in a 50 mL conical tube.
3. Measure 10 mL of acetonitrile and add to the tube.
4. Mix vigorously for 5 minutes.
5. Centrifuge for 5 minutes at 3,000 x g.
6. Transfer 1 mL of the supernatant to a clean container.
7. Dry the sample with a gentle nitrogen stream.
8. Measure 1 mL of Sample Buffer 2 and add to the dried sample.
9. Mix vigorously and use in the assay.

Honey: (Dilution Factor: 20)

1. Weigh 1 gram of sample in a glass, screw cap bottle.
2. Measure 19 mL of Sample Buffer 3 and add to the bottle.
3. Place the bottle in an ultrasonic water bath for 5 minutes.
4. Mix vigorously for 2 minutes.
5. Mix vigorously again prior to use in the assay.

Assay Procedure

1. Allow kit components and the sample extract(s) to reach room temperature prior to running the test.
2. Place the appropriate number of test wells into a holder. Be sure to re-seal unused test wells in the zip-lock bag with the desiccant to limit exposure to moisture.
3. Dispense **50 μ L of Enzyme Conjugate** into each well.
4. Dispense **50 μ L of Calibrators and Sample Extract(s)** into the appropriate well. Be sure to use a clean pipette tip for each solution to avoid cross contamination.
5. Dispense **50 μ L of Antibody** into each well.
6. Gently shake the wells for 30 seconds using a back-and-forth motion and incubate for **30 minutes** at room temperature.
7. Decant the contents of the wells into an appropriate waste container. Fill the wells to overflowing with laboratory quality distilled or deionized water and then decant. Repeat this wash step four times for a total of five washes. Following the last wash, tap the inverted wells onto absorbent paper to remove excess wash solution.
8. Dispense **100 μ L of Substrate** into each well.
9. Gently shake the wells for 30 seconds using a back-and-forth motion and incubate for **30 minutes** at room temperature.
10. Dispense **100 μ L of Stop Solution** into each well in the same order of addition as the Substrate.
11. Gently shake the wells for 30 seconds using a back-and-forth motion.
12. Carefully wipe the optical surface with a soft, lint-free wipe. Measure and record the absorbance (Optical Density; OD) of each well at 450 nm using a plate or strip reader within 10 minutes of stopping the assay. If the reader has dual wavelength capability, read at 450 nm minus 605 or 650 nm.

Result Interpretation

Semi-Quantitative Interpretation: Semi-quantitative results can be derived by simple comparison of the sample absorbances to the absorbance of the calibrators:

- Samples with a lower absorbance (less color) than a calibrator have a concentration of E/C greater than the concentration of the calibrator.
- Samples with a higher absorbance (more color) than a calibrator have a concentration less than the concentration of the calibrator.

Quantitative Interpretation: It is preferred for quantitative results to be determined using commercially available software for ELISA evaluation using a 4-parameter curve fit. Alternatively, a semi-log curve fit can be used if 4-parameter software is not available. A spreadsheet that will perform the curve fit and sample concentration calculations is available upon request. Please contact Beacon for further details.

- The concentration of E/C in a sample is determined by comparing the average sample absorbance to the standard curve. This value must then be multiplied by the dilution factor used.
- Samples with absorbances lower than the highest calibrator contain a concentration of E/C too high for quantification. Further dilute the sample extract to fit into the standard curve and retest along with the calibrators. Results must then be multiplied by the dilution factor used.
- Samples with E/C absorbances greater than the lowest calibrator or less than the highest calibrator must be reported as < 0.1 ppb or > 5 ppb, respectively.

Technical Assistance

For questions regarding this kit or for additional information about Beacon products, contact us.

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Safety

Stop Solution is 1N hydrochloric acid. Handle with care. To receive complete safety information on this product, contact Beacon Analytical Systems, Inc., and request Safety Data Sheets.

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